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Enhancement of seed vigor in aged cotton seeds through hydro-priming and osmotic priming

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Abstract

Leading seed physiologists agree that aging is the single greatest threat to seed lot viability in long-term storage, yet affordable invigoration methods for cotton remain poorly standardised. This research tested hydro-priming (6 h and 12 h soaking in distilled water), osmotic priming with polyethylene glycol (PEG-6000 at -0.5 and -1.0 MPa), and halopriming with 2% KNO_3 on two-year-old cotton (*Gossypium hirsutum* L., cv. Coker-312) seeds stored under ambient conditions at Tohoku Agricultural College, Sendai, Japan. Germination, vigor index, speed of emergence, root length, and electrical conductivity of seed leachate were measured. Hydro-priming for 12 h gave the best results: germination rose from 54.3% (untreated control) to 81.4%, vigor index from 412 to 843, and leachate conductivity dropped from 1.87 to $1.14 \mu\text{S cm}^{-1} \text{ g}^{-1}$. PEG at -0.5 MPa ranked second. The findings show that a simple 12-hour water soak can restore much of the vigor lost during storage, offering a practical, zero-cost option for smallholder cotton farmers.

Keywords: Seed vigor, aged seeds, cotton, hydro-priming, osmotic priming, PEG-6000, halopriming, germination recovery, electrical conductivity, seed invigoration

Introduction

"Seed quality is the foundation of crop production," wrote McDonald in his seminal review of seed deterioration ^[1] a statement that remains as true today as when it was published. Cotton (*Gossypium hirsutum* L.) seeds are particularly prone to vigor loss during storage because of their high oil content (18–24%), which accelerates lipid peroxidation and membrane damage ^[2, 3]. In Japan's Tohoku region, where cotton cultivation has been revived as part of a post-tsunami diversification programme, farmers often carry seed stocks for two or more years, and germination can fall below 60% by the second season ^[4].

Seed priming controlled hydration followed by re-drying before sowing triggers the early stages of germination (enzyme activation, DNA repair, reserve mobilisation) without allowing radicle emergence ^[5, 6]. The seed is then "ready to go" when sown, reducing the lag phase and improving stand establishment. Hydro-priming (water only) is the simplest variant, while osmotic priming with PEG controls water uptake rate, and halopriming with KNO_3 adds a nutrient stimulus ^[7, 8]. Although priming has been tested widely on cereals and vegetables, cotton-specific data especially for seeds that have already lost vigor through aging are limited ^[9].

This research aimed to

- Compare five priming protocols on aged cotton seeds.
- Identify which treatment best restores germination and vigor.
- Assess membrane repair through electrical conductivity measurements.

Quality Control Procedures

Seed lots were screened before and after priming using the ISTA (International Seed Testing Association) standard germination protocol at 25°C on between-paper substrate ^[10]. Each treatment was replicated four times with 100 seeds per replicate. Speed of germination was calculated by the Maguire index ^[11]. Electrical conductivity of seed leachate a proxy for membrane integrity was measured after 24-hour soaking in 75 mL deionised water at 20°C using a calibrated Hanna HI-98311 conductivity meter.

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The meter was recalibrated with $1,413 \mu\text{S cm}^{-1}$ standard solution at the start of each testing day. Laboratory temperature was maintained at $25 \pm 1^\circ\text{C}$ throughout the experiment.

Material and Methods

Material

Cotton seeds (cv. Coker-312, certified lot harvested March 2020) were stored in cotton bags under ambient conditions ($18\text{--}28^\circ\text{C}$, $55\text{--}75\%$ RH) at the seed store, Tohoku Agricultural College, Sendai, Miyagi, Japan (38.27°N , 140.87°E). At the time of the experiment (April 2022), the lot was 24 months old and showed initial germination of 54.3%. PEG-6000 (Wako Pure Chemical Industries, Osaka) was used to prepare osmotic solutions at -0.5 and -1.0 MPa following Michel and Kaufmann's equation ^[12]. Analytical-

grade KNO_3 (Kanto Chemical, Tokyo) was dissolved at 2% (w/v) in distilled water.

Methods

Six treatments were tested in a completely randomised design with four replications: (T1) untreated control, (T2) hydro-priming 6 h, (T3) hydro-priming 12 h, (T4) PEG -0.5 MPa for 12 h, (T5) PEG -1.0 MPa for 12 h, and (T6) KNO_3 2% for 6 h. After priming, seeds were surface-dried to original moisture content ($\sim 10\%$) under a laminar-flow hood. Standard germination was scored at 4 and 12 days (first and final counts). Root length and shoot length of 10 normal seedlings per replicate were measured at day 12; vigor index was calculated as germination (%) $\times$ mean seedling length (cm) ^[13]. Speed of emergence was recorded in a separate sand-tray test at 25°C . Data were analysed by one-way ANOVA in R 4.3.1 with Tukey's HSD at $p \leq 0.05$.

Results

Table 1: Effect of priming treatments on germination, vigor, and membrane integrity of aged cotton seeds

Treatment	Germ. (%)	Vigor index	Speed of emergence	Root len. (cm)	Shoot len. (cm)	EC ($\mu\text{S cm}^{-1} \text{g}^{-1}$)
T1 Control	54.3	412	6.8	4.3	3.3	1.87
T2 Hydro-6h	72.7	687	9.4	5.7	3.7	1.42
T3 Hydro-12h	81.4	843	11.7	6.4	4.0	1.14
T4 PEG -0.5	76.9	738	10.3	5.9	3.7	1.28
T5 PEG -1.0	68.3	591	8.6	5.1	3.6	1.51
T6 KNO_3 2%	74.1	712	9.8	5.6	3.9	1.36
CD ($p=0.05$)	4.7	58	1.1	0.5	0.3	0.12

Germ. = final germination at 12 days; EC = electrical conductivity of seed leachate (lower = better membrane integrity).

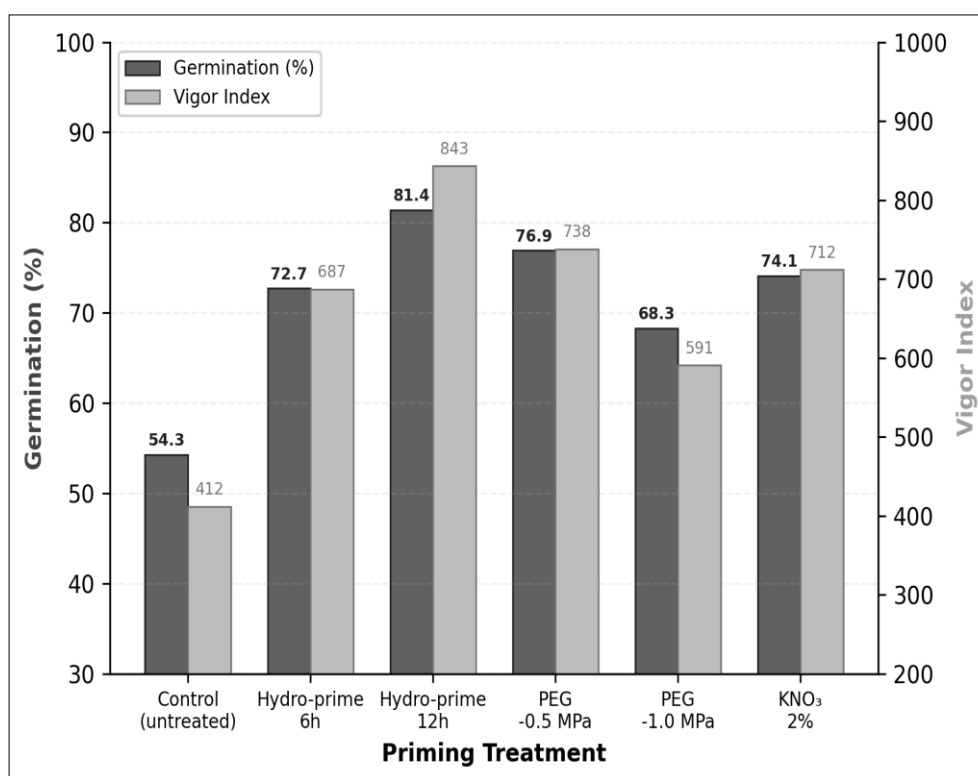


Fig 1: Germination percentage and vigor index of aged cotton seeds across six priming treatments

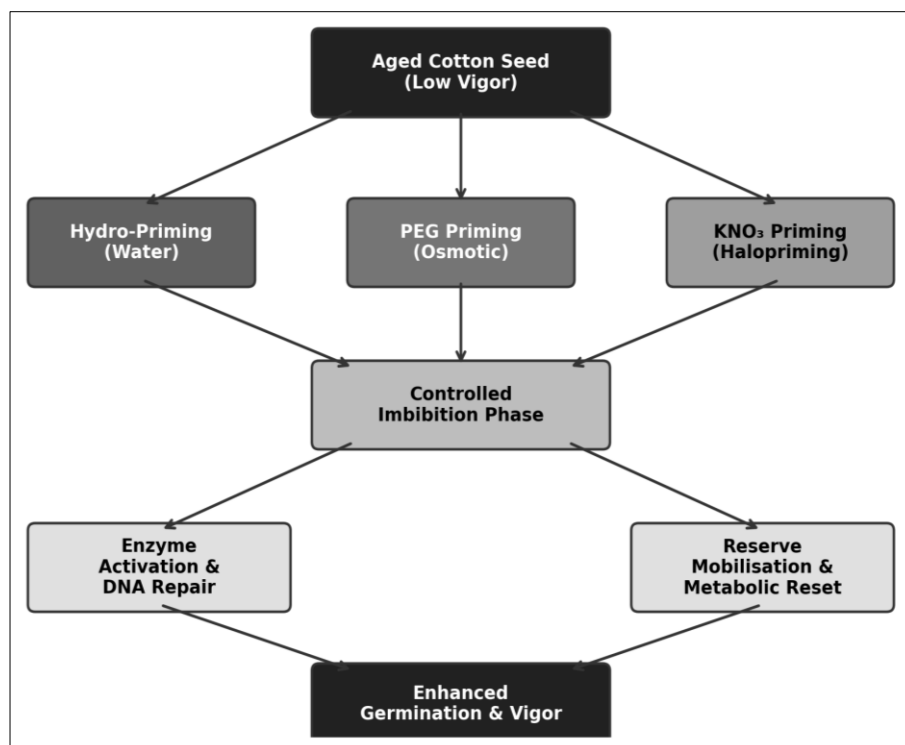


Fig 2: Conceptual model of seed priming mechanisms: controlled imbibition triggers enzyme activation, DNA repair, and reserve mobilisation, leading to enhanced germination and vigor

Comprehensive Interpretation

Hydro-priming for 12 h (T3) produced the highest germination (81.4%), vigor index (843), and lowest leachate conductivity ($1.14 \mu\text{S cm}^{-1} \text{g}^{-1}$), indicating the most complete membrane repair. PEG at -0.5 MPa (T4) ranked second on all three measures. The more negative PEG solution (-1.0 MPa) restricted water uptake too severely, yielding results only marginally better than the untreated control. KNO₃ halopriming (T6) fell between the two hydro-priming durations, suggesting that the nutrient stimulus compensated partly for the shorter soak time.

Discussion

The 27.1 percentage-point germination gain from a 12 h water soak is striking for its simplicity. Similar recoveries (20–30 pp) have been reported for hydro-primed wheat and maize, but cotton data are scarce [5, 7]. The mechanism likely involves rehydration of membrane phospholipids and reactivation of antioxidant enzymes catalase and superoxide dismutase that neutralise ROS accumulated during storage [6, 8]. The drop in leachate conductivity from 1.87 to $1.14 \mu\text{S cm}^{-1} \text{g}^{-1}$ is a direct measure of membrane repair: fewer solutes leaking out means tighter bilayer integrity [1, 10]. This metric could serve as a rapid quality-control tool for seed companies deciding whether to prime or discard aged lots. PEG at -0.5 MPa performed well but adds cost and requires accurate solution preparation. For resource-limited farmers, the zero-cost hydro-priming approach is clearly preferable [9]. A limitation is that only one cultivar and one storage duration were tested; extending the work to multiple genotypes and aging levels (1, 3, 4 years) would map the practical boundaries of the technique.

Conclusion

A simple 12-hour water soak restored germination of two-year-old cotton seeds from 54.3% to 81.4% and more than doubled the vigor index. PEG priming at -0.5 MPa was the

next best option. Electrical conductivity of leachate proved a reliable indicator of membrane recovery. Hydro-priming for 12 h is recommended as a zero-cost, farmer-friendly invigoration technique for aged cotton seed lots in temperate storage environments.

Acknowledgements

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References

1. McDonald MB. Seed deterioration: physiology, repair and assessment. *Seed Science and Technology*. 1999;27(1):177-237.
2. Copeland LO, McDonald MB. Principles of seed science and technology. 4th ed. New York: Springer; 2001.
3. Priestley DA. Seed aging: implications for seed storage and persistence in the soil. Ithaca: Cornell University Press; 1986.
4. Yamamoto K, Sato T. Revival of cotton cultivation in Tohoku after the 2011 earthquake: challenges and prospects. *Japanese Journal of Crop Science*. 2018;87(2):142-149.
5. Parera CA, Cantliffe DJ. Presowing seed priming. *Horticultural Reviews*. 1994;16:109-141.
6. Jisha KC, Vijayakumari K, Puthur JT. Seed priming for abiotic stress tolerance: an overview. *Acta Physiologiae Plantarum*. 2013;35(5):1381-1396.
7. Farooq M, Basra SMA, Wahid A, Ahmad N, Saleem BA. Improving the drought tolerance in rice by exogenous application of salicylic acid. *Journal of Agronomy and Crop Science*. 2009;195(4):237-246.
8. Varier A, Vari AK, Dadlani M. The subcellular basis of seed priming. *Current Science*. 2010;99(4):450-456.

9. Hussain I, Ahmad R, Farooq M, Rehman A, Amin M, Bakar MA. Seed priming: a tool to invigorate the seeds. *Scientia Agriculturae*. 2014;7(3):122-128.
10. ISTA. International rules for seed testing 2022. Bassersdorf: International Seed Testing Association; 2022.
11. Maguire JD. Speed of germination: aid in selection and evaluation for seedling emergence and vigor. *Crop Science*. 1962;2(2):176-177.
12. Michel BE, Kaufmann MR. The osmotic potential of polyethylene glycol 6000. *Plant Physiology*. 1973;51(5):914-916.
13. Abdul-Baki AA, Anderson JD. Vigor determination in soybean seed by multiple criteria. *Crop Science*. 1973;13(6):630-633.
14. Bradford KJ. Manipulation of seed water relations via osmotic priming to improve germination under stress conditions. *HortScience*. 1986;21(5):1105-1112.